

# VetLine Coxiella Phase 2

**ELISA**

**RUO**

**For Research Use Only**

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Product Number:      COX2VT0600 (96 Determinations)

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## ENGLISH

### 1. INTRODUCTION

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### 2. INTENDED USE

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### 3. PRINCIPLE OF THE ASSAY

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The qualitative immunoenzymatic determination of specific antibodies is based on the ELISA (Enzyme-linked Immunosorbent Assay) technique.

Microtiterplates are coated with specific antigens to bind corresponding antibodies of the sample. After washing the wells to remove all unbound sample material a horseradish peroxidase (HRP) labelled conjugate is added. This conjugate binds to the captured antibodies. In a second washing step unbound conjugate is removed. The immune complex formed by the bound conjugate is visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product.

The intensity of this product is proportional to the amount of specific antibodies in the sample. Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450/620 nm is read using an ELISA Microtiterplate reader.

### 4. MATERIALS

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#### 4.1. Reagents supplied

- **Microtiterplate:** 12 breakapart 8-well snap-off strips coated with Coxiella antigens; in resealable aluminium foil.
- **Sample Dilution Buffer:** 1 bottle containing 100 mL of phosphate buffer (10 mM) for sample dilution; pH 7.2 ± 0.2; coloured yellow; ready to use; white cap; ≤ 0.0015 % (v/v) CMIT/MIT (3:1).
- **Stop Solution:** 1 bottle containing 15 mL sulphuric acid, 0.2 mol/L; ready to use; red cap.
- **Washing Buffer (20x conc.):** 1 bottle containing 50 mL of a 20-fold concentrated phosphate buffer (0.2 M), pH 7.2 ± 0.2, for washing the wells; white cap.
- **Conjugate:** 1 bottle containing 20 mL of peroxidase labelled Protein A/G; coloured yellow, ready to use; white cap; ≤ 0.02 % (v/v) MIT.
- **TMB Substrate Solution:** 1 bottle containing 15 mL 3,3',5,5'-tetramethylbenzidine (TMB), < 0.1 %; ready to use; yellow cap.
- **Positive Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; red cap; ≤ 0.02 % (v/v) MIT.
- **Cut-off Control:** 1 vial containing 3 mL control; coloured yellow; ready to use; green cap; ≤ 0.02 % (v/v) MIT.
- **Negative Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; blue cap; ≤ 0.0015 % (v/v) CMIT/MIT (3:1).

For hazard and precautionary statements see 12.1

For potential hazardous substances please check the safety data sheet.

#### 4.2. Materials supplied

- 1 Cover foil
- 1 Instruction for use (IFU)
- 1 Plate layout

#### 4.3. Materials and Equipment needed

- ELISA Microtiterplate reader, equipped for the measurement of absorbance at 450/620 nm
- Incubator 37 °C
- Manual or automatic equipment for rinsing Microtiterplate wells
- Pipettes to deliver volumes between 10 and 1000 µL
- Vortex tube mixer
- Distilled water
- Disposable tubes

### 5. STABILITY AND STORAGE

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Store the kit at 2...8 °C. The opened reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

### 6. REAGENT PREPARATION

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It is very important to bring all reagents and samples to room temperature (20...25 °C) and mix them before starting the test run!

#### 6.1. Microtiterplate

The break-apart snap-off strips are coated with Coxiella antigens. Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8 °C.

## 6.2. Washing Buffer (20x conc.)

Dilute Washing Buffer 1 + 19; e. g. 10 mL Washing Buffer + 190 mL distilled water. The diluted buffer is stable for 5 days at room temperature (20...25 °C). In case crystals appear in the concentrate, warm up the solution to 37 °C e.g. in a water bath. Mix well before dilution.

## 6.3. TMB Substrate Solution

The reagent is ready to use and has to be stored at 2...8 °C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away.

## 7. SAMPLE COLLECTION AND PREPARATION

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Use veterinary serum or milk samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

- Pooled serum: A maximum of five samples can be pooled
- Milk samples: Whole milk samples can be used after centrifugation for 15 min at 2.000 x g or 2 min at 10.000 x g  
The sample should be drawn from below the cream layer

### 7.1. Sample Dilution

#### Single Serum Sample:

Before assaying all serum samples should be diluted 1+100 with Sample Dilution Buffer. Dispense 10 µL sample and 1 mL Sample Dilution Buffer into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

#### Pooled Serum Sample:

Before assaying up to 5 serum samples should be diluted with Sample Dilution Buffer in the same tube.

Dispense 10 µL of each sample in the same tube with 1 mL Sample Dilution Buffer to obtain max 50 µL serum in 1 mL Sample Dilution Buffer and thoroughly mix with a Vortex.

#### Milk Sample:

Before assaying all milk samples should be diluted 1+10 with Sample Dilution Buffer. Dispense 100 µL milk and 1 mL Sample Dilution Buffer into tubes to obtain a 1+10 dilution and thoroughly mix with a Vortex.

Optimal milk dilution might differ from region to region and between animal species. It may range from undiluted to 1+20 and has to be determined by each lab by itself. The described 1+10 dilution has to be considered as a starting point for evaluation.

## 8. ASSAY PROCEDURE

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Please read the instruction for use carefully **before** performing the assay. Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend increasing the washing steps from three up to five and the volume of Washing Buffer from 300 µL to 350 µL to avoid washing effects. Pay attention to chapter 12. Prior to commencing the assay, the distribution and identification plan for all samples and standards/controls (duplicates recommended) should be carefully established on the plate layout supplied in the kit. Select the required number of microtiter strips or wells and insert them into the holder.

Perform all assay steps in the order given and without any delays.

A clean, disposable tip should be used for dispensing each standard/control and sample.

Adjust the incubator to 37 ± 1 °C.

1. Dispense 100 µL standards/controls and diluted samples into their respective wells. Leave well A1 for the Substrate Blank.
2. Cover wells with the foil supplied in the kit.
3. **Incubate for 1 hour ± 5 min at 37 ± 1 °C.**
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300 µL of Washing Buffer. Avoid overflows from the reaction wells. The interval between washing and aspiration should be > 5 sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!  
Note: Washing is important! Insufficient washing results in poor precision and false results.
5. Dispense 100 µL Conjugate into all wells except for the Substrate Blank well A1.
6. **Incubate for 30 min at room temperature (20...25 °C).** Do not expose to direct sunlight.
7. Repeat step 4.
8. Dispense 100 µL TMB Substrate Solution into all wells.
9. **Incubate for exactly 15 min at room temperature (20...25 °C) in the dark.** A blue colour occurs due to an enzymatic reaction.
10. Dispense 100 µL Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution, thereby a colour change from blue to yellow occurs.
11. Measure the absorbance at 450/620 nm within 30 min after addition of the Stop Solution.

## 8.1. Measurement

Adjust the ELISA Microtiterplate reader to **zero** using the **Substrate Blank**.

If - due to technical reasons - the ELISA Microtiterplate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results!

**Measure the absorbance** of all wells at **450 nm** and record the absorbance values for each standard/control and sample in the plate layout.

Bichromatic measurement using a reference wavelength of 620 nm is recommended.

Where applicable calculate the **mean absorbance values** of all duplicates.

## 9. RESULTS

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### 9.1. Run Validation Criteria

In order for an assay to be considered valid, the following criteria must be met:

- **Substrate Blank:** Absorbance value < **0.100**
- **Negative Control:** Absorbance value < **0.200** and < **Cut-off**
- **Cut-off Control:** Absorbance value **0.150 – 1.300**
- **Positive Control:** Absorbance value > **Cut-off**

If these criteria are not met, the test is not valid and must be repeated.

### 9.2. Calculation of Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations.

Example: Absorbance value Cut-off Control 0.44 + absorbance value Cut-off control 0.42 = 0.86 / 2 = 0.43  
Cut-off = 0.43

#### 9.2.1. Results in Units [NTU]

$$\frac{\text{Sample (mean) absorbance value} \times 10}{\text{Cut-off}} = [\text{NovaTec Units} = \text{NTU}]$$

Example: 
$$\frac{1.591 \times 10}{0.43} = 37 \text{ NTU}$$

### 9.3. Interpretation of Results

Cut-off	10 NTU
Positive	> 11 NTU
Equivocal	9 – 11 NTU
Negative	< 9 NTU

## 10. SPECIFIC PERFORMANCE CHARACTERISTICS

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For further information about the specific performance characteristics please contact NovaTec Immundiagnostica GmbH.

### 10.1. Precision

### 10.2. Diagnostic Specificity

### 10.3. Diagnostic Sensitivity

### 10.4. Interferences

Interferences with hemolytic, lipemic or icteric samples are not observed up to a concentration of 10 mg/mL hemoglobin, 5 mg/mL triglycerides and 0.5 mg/mL bilirubin.

### 10.5. Cross Reactivity

## 11. LIMITATIONS OF THE PROCEDURE

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Bacterial contamination or repeated freeze-thaw cycles of the sample may affect the absorbance values.

## 12. PRECAUTIONS AND WARNINGS

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- **For research use only!**
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- All components of human origin used for the production of these reagents have been tested for anti-HIV antibodies, anti-HCV antibodies and HBsAg and have been found to be non-reactive.
- Do not interchange reagents or strips of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette samples and dispense reagents without splashing accurately into the wells.
- The ELISA is only designed for qualified personnel who are familiar with good laboratory practice.

### 12.1. Safety note for reagents containing hazardous substances

Reagents may contain CMIT/MIT (3:1) or MIT (refer to 4.1)

Therefore, the following hazard and precautionary statements apply.

**Warning**



H317	May cause an allergic skin reaction.
P261	Avoid breathing spray
P280	Wear protective gloves/protective clothing.
P302+P352	IF ON SKIN: Wash with plenty of soap and water.
P333+P313	If skin irritation or rash occurs: Get medical advice/attention.
P362+P364	Take off contaminated and Wash it before reuse.

Further information can be found in the safety data sheet.

### 12.2. Disposal Considerations

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

## 13. ORDERING INFORMATION

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
Prod. No.:            COX2VT0600        VetLine Coxiella Phase 2 ELISA (96 Determinations)

**BIBLIOGRAPHY / LITERATUR / BIBLIOGRAPHIE / BIBLIOGRAFIA / BIBLIOGRAFÍA/  
BLIBIOGRAFIA**

**ABBREVIATIONS / ABKÜRZUNGEN / ABRÉVIATIONS / ABBREVIAZIONI / ABREVIACIONES /  
ABREVIATURAS**

<b>CMIT</b>	5-chloro-2-methyl-4-isothiazolin-3-one
<b>MIT</b>	2-methyl-2H-isothiazol-3-one

**SYMBOLS KEY / SYMBOLSCHLÜSSEL / EXPLICATION DES SYMBOLES / LEGENDA / SIMBOLOS / TABELA DE SIMBOLOS**

	Manufactured by / Hergestellt von / Fabriqué par / Prodotto da / Fabricado por / Fabricado por
<b>LOT</b>	Lot Number / Chargenbezeichnung / Numéro de lot / Lotto / Número de lote / Número de lote
	Expiration Date / Verfallsdatum / Date de péremption / Scadenza / Fecha de caducidad / Data de Validade
	Storage Temperature / Lagertemperatur / Température de conservation / Temperatura di conservazione / Temperatura de almacenamiento / Temperatura de Armazenamento
<b>RUO</b>	For research use only/ Nur für Forschungszwecke / Destiné à la recherche uniquement/ Solo per scopi di ricerca/ Uso exclusivo en investigación / Apenas para fins de pesquisa
<b>REF</b>	Catalogue Number / Katalog Nummer / Référence du catalogue / Numero di codice / Número de Catálogo / Número de Catálogo
	Consult Instructions for Use / Arbeitsanleitung beachten / Consulter la notice d'utilisation / Consultare le istruzioni per l'uso / Consulte las Instrucciones de Uso / Consultar as Instruções de Utilização
<b>MTP</b>	Microtiterplate / Mikrotiterplatte / Plaque de Microtitrage / Piastre di Microtitolazione / Placa de Microtitulación / Placa de Microtitulação
<b>CONJL</b>	Conjugate / Konjugat / Conjugué / Coniugato / Conjugado / Conjugado
<b>CONTROL -</b>	Negative Control / Negativkontrolle / Contrôle Négatif / Controllo Negativo / Control Negativo / Controle Negativo
<b>CONTROL +</b>	Positive Control / Positivkontrolle / Contrôle Positif / Controllo Positivo / Control Positivo / Controle Positivo
<b>CUT OFF</b>	Cut-off Control / Cut-off Kontrolle / Contrôle Cut-off / Controllo Cut-off / Control Cut-off / Controle Cut-off
<b>DIL</b>	Sample Dilution Buffer / Probenverdünnungspuffer / Tampon de Dilution d'Échantillon / Tampone di Diluizione del Campione / Tampón de Dilución de Muestras / Tampão de Diluição de Amostra
<b>SOLN STOP</b>	Stop Solution / Stopplösung / Solution d'Arrêt / Soluzione Bloccante / Solución de Parada / Solução de Bloqueio
<b>SUB TMB</b>	TMB Substrate Solution / TMB-Substratlösung / Solution de Substrat TMB / Soluzione Substrato TMB / Solución de Sustrato de TMB / Solução Substrato TMB
<b>WASH BUF 20x</b>	Washing Buffer 20x concentrated / Waschpuffer 20x konzentriert / Tampon de Lavage concentré 20 x / Tampone di Lavaggio concentrazione x20 / Tampón de Lavado concentrado x20 / Tampão de Lavagem concentrada 20x
	Contains sufficient for "n" tests / Ausreichend für "n" Tests / Contenu suffisant pour "n" tests / Contenido suficiente per "n" saggi / Contenido suficiente para "n" tests / Conteúdo suficiente para "n" testes

**SUMMARY OF TEST PROCEDURE / KURZANLEITUNG TESTDURCHFÜHRUNG / RÉSUMÉ DE LA PROCEDURE DE TEST / SCHEMA DELLA PROCEDURA / RESUMEN DE LA TÉCNICA / RESUMO DO PROCEDIMENTO DE TESTE**

# SCHEME OF THE ASSAY

VetLine Coxiella Phase 2 ELISA

## Test Preparation

Prepare reagents and samples as described.  
 Establish the distribution and identification plan for all samples and standards/controls on the plate layout supplied in the kit.  
 Select the required number of microtiter strips or wells and insert them into the holder.

## Assay Procedure

	Substrate Blank (A1)	Negative Control	Cut-off Control	Positive Control	Sample <small>(dilution: refer to point 7.1)</small>
Negative Control	-	100 µL	-	-	-
Cut-off Control	-	-	100 µL	-	-
Positive Control	-	-	-	100 µL	-
Sample <small>(dilution: refer to point 7.1)</small>	-	-	-	-	100 µL
Cover wells with foil supplied in the kit <b>Incubate for 1 h at 37±1 °C</b> Wash each well three times with 300 µl of Washing Buffer					
Conjugate	-	100 µL	100 µL	100 µL	100 µL
<b>Incubate for 30 min at room temperature (20...25 °C)</b> Do not expose to direct sunlight Wash each well three times with 300 µL of Washing Buffer					
TMB Substrate Solution	100 µL	100 µL	100 µL	100 µL	100 µL
<b>Incubate for exactly 15 min at room temperature (20...25 °C) in the dark</b>					
Stop Solution	100 µL	100 µL	100 µL	100 µL	100 µL
Photometric measurement at 450 nm (reference wavelength: 620 nm)					



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