

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/287505647>

A cross-sectional study of the impact of regular use of insecticides in dogs on Canine Leishmaniosis seroprevalence in southeast Spain

ARTICLE *in* PREVENTIVE VETERINARY MEDICINE · DECEMBER 2015

Impact Factor: 2.17 · DOI: 10.1016/j.prevetmed.2015.12.009

READS

14

7 AUTHORS, INCLUDING:



[Pedro Perez Cutillas](#)

University of Murcia

37 PUBLICATIONS 3 CITATIONS

[SEE PROFILE](#)

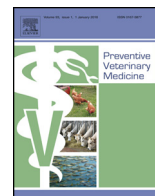


[Eduardo Berriatua](#)

University of Murcia

60 PUBLICATIONS 1,023 CITATIONS

[SEE PROFILE](#)



A cross-sectional study of the impact of regular use of insecticides in dogs on Canine Leishmaniosis seroprevalence in southeast Spain



E. Goyena^a, P. Pérez-Cutillas^b, L. Chitimia^a, J. Risueño^a, J.D. García-Martínez^c, L.J. Bernal^c, E. Berriatua^{a,*}

^a Departamento de Sanidad Animal, Facultad de Veterinaria, Regional Campus of International Excellence “Campus Mare Nostrum”, Universidad de Murcia, 30100 Murcia, Spain

^b Centro de Edafología y Biología Aplicada del Segura, Consejo Superior de Investigaciones Científicas (CEBAS-CSIC), 30100 Murcia, Spain

^c Departamento de Medicina y Cirugía Animal, Facultad de Veterinaria, Regional Campus of International Excellence “Campus Mare Nostrum”, Universidad de Murcia, 30100 Murcia, Spain

ARTICLE INFO

Article history:

Received 31 August 2015

Received in revised form

27 November 2015

Accepted 15 December 2015

Keywords:

Canine Leishmaniosis

Insecticides

Seroprevalence

Murcia

Spain

ABSTRACT

The relationship between Canine Leishmaniosis (CanL) seroprevalence and regular use of topical insecticides was investigated in 800 pet dogs with no visible signs of CanL in Murcia, southeast Spain in 2011. Dogs were clients to 17 veterinary practices and were analyzed for *Leishmania infantum* antibodies in blood plasma using two commercial ELISAs (Ingezim, Ingenasa[®], Spain; Leishcan, Hipra[®], Spain). Owners were interviewed to gather data on dog related variables. They included date of birth, home address and frequency, duration and timing of insecticide treatments used to prevent ectoparasite infestations. The dog's residence was georeferenced and environmental data potentially associated with the dog's risk of *L. infantum* infection was obtained. A mixed logistic regression model was then developed to analyze the relationship between the dog's serological status and insecticidal treatment adjusted for demographic and environmental variables. Overall, CanL seroprevalence (95% confidence limits) was 18% (16–21%) including 11% in dogs not using insecticide treatments ($n=60$) and 19% in those using them ($n=740$) ($p>0.05$). At least 16 different insecticide products were used and 73%, 26% and 1% of dogs received 1, 2 and 3 products a year. The most frequent commercial brands used and the only ones in the market claiming anti-sandfly activity, were Scalibor collars (deltamethrin 40 mg/g; MSD[®]), Advantix pipettes (permethrin 500 mg/ml and imidacloprid 100 mg/ml; Bayer[®]) and Exspot spot-on pipettes (permethrin 715 mg/ml; MSD[®]). Seroprevalence was 9%, 16%, 20%, 22% and 25% for dogs with Scalibor collars plus Advantix pipettes, Scalibor collars plus ExSpot pipettes, Advantix pipettes alone, Scalibor collars alone and Exspot pipettes alone, respectively. The multivariable model confirmed a significant reduction in the risk of *Leishmania* spp. seropositivity in dogs using the Scalibor and Advantix combination compared to those using either product alone and provided evidence of greatly increased risk of CanL in rural areas situated at 300–500 m altitude and average March–July temperatures of 18.6–19 °C. The study highlights the difficulty in controlling CanL infection by means of insecticide use alone and that it could be improved by using the Scalibor and Advantix combination and identifying and targeting specific geographical areas.

© 2015 Elsevier B.V. All rights reserved.

1. Introduction

Canine Leishmaniosis (CanL) caused by *Leishmania infantum* is a major vector borne zoonotic infection in Mediterranean Spain causing mortality and a spectrum of severe clinical outcomes (Baneth et al., 2008). Its impact in endemic areas is further exacerbated by the difficulty in preventing infection (Otranto and Dantas-Torres,

2013). The efficacy of available vaccines is still under assessment and treatment of infected animals doesn't achieve parasitological cure. Moreover, it's not implemented on asymptotically infected animals, which represent 50–80% of dogs living in some endemic areas in Spain (Solano-Gallego et al., 2001; Chitimia et al., 2011). In European countries CanL control efforts primarily focusses on preventing phlebotomine sandfly vector biting by using insecticides with residual activity in the dog (Otranto and Dantas-Torres, 2013).

When this study was carried out there were no leishmania vaccines in Spain and three commercial insecticide products specifically marketed to prevent phlebotomine sandfly vector

* Corresponding author. Fax: +34 868884147.
E-mail address: berriatu@um.es (E. Berriatua).

infection were available: Scalibor insecticide-impregnated collars (deltamethrin 40 mg/g; MSD®), Advantix topical (spot-on) pipettes (permethrin 500 mg/ml and imidacloprid 100 mg/ml; Bayer®) and Exspot spot-on pipettes (permethrin 715 mg/ml; MSD®). In laboratory conditions all three products have a sandfly repellent (blood feeding avoidance on a treated dog) efficacy greater than 90% and an insecticidal efficacy (additional mortality of fed and unfed sandflies exposed to a treated dog) of 30–99%, during the first three weeks after application (Killick-Kendrick et al., 1997; Mencke et al., 2003; Miró et al., 2007; Molina et al., 2012). Controlled field trials in pets and kennel dogs in places with a high CanL prevalence corroborate that both Scalibor collars and Advantix and Exspot pipettes can be efficacious at preventing infection in dogs. However, the reduction in annual CanL seroconversion ranged from 50 to 100% depending on the year and the product used (Maroli et al., 2001; Foglia Manzillo et al., 2006; Otranto et al., 2007, 2010; Ferroglio et al., 2008). The residual activity of these products is time dependent, lasting 3 to 4 weeks for pipettes and around six months for the collar however, collar losses can be a major problem (Foglia Manzillo et al., 2006). Other factors conditioning the practicality of collars as a tool for controlling CanL is the percentage of the dog population collared and the infection incidence (the greater, the more useful collars are) (Maroli et al., 2001; Reithinger et al., 2004).

There is however a lack of information on the performance of dog insecticides to prevent CanL infection in the wide dog population in endemic areas in uncontrolled field conditions. Such studies may be hampered by the difficulty in comparing the risk of infection in heterogeneous dog populations with widely variable exposure to infected sandflies. In the present study we employed multivariable regression modeling to investigate the relationship between regular use of anti-sandfly insecticide products and dogs CanL serological antibody status in endemic Murcia Region, to control for critical dog's demographic and residential environmental variables. CanL seroprevalence in Murcia was recently estimated at 7% for dogs from periurban residential areas (Chitimia et al., 2011).

2. Materials and methods

2.1. Study population and data collection

The study was carried out on up to 800 pet dogs aged 6 months-old or older, with no visible clinical signs of CanL, clients to 17 veterinary practices across Murcia Region, in November 2011. This is a traditionally endemic southern Mediterranean area (de Ybáñez et al., 2009) situated 38°N and 1°50'W, ranging 11300 km², with approximately 1470000 inhabitants. There is no recent information on sandfly abundance in the region. Studies in the 1980s showed that *Phlebotomus perniciosus* was the main *L. infantum* vector in Murcia with peaks of maximum abundance in June and September and no activity recorded from mid-November until April (Martínez Ortega and Conesa Gallego, 1987).

Dogs selected attended the veterinary practice to participate in the study as they were offered a CanL serological analysis free of charge, or for a consultation unrelated to CanL. Upon selection, dogs were clinically examined to discard animals with visible CanL signs (Baneth et al., 2008) and were then blood sampled and weighed. Owners were asked to answer a brief questionnaire about the dog's date of birth, gender, breed, residential home address and type, presence of people and animals in their home, sleeping place and insecticide usage in the animal (Tables 1–3). The specific questions concerning insecticide use were: (i) do you use insecticides in the dog on a regular basis to protect it against insects, mites and ticks? if you do, (ii) which insecticide brand or brands do you use on your

Table 1

Canine Leishmaniosis seroprevalence (95% confidence intervals) according to dog's signalment and body weight. A cross-sectional study in dogs from Murcia, southeast Spain in autumn 2011.

Variable	Level	N	ELISA			P value
			% Positive	95–	95+	
Gender	Female	426	15	11	18	0.0041
	Male	375	23	18	27	
Age (years)	0.5–1	172	9	5	14	0.0484
	2	112	16	9	23	
	3	103	16	9	23	
	4	72	21	11	30	
	5	57	19	9	30	
	6	69	22	12	31	
	7	44	25	12	38	
	8	25	24	7	41	
	9	33	27	12	42	
	10	27	30	12	47	
	>10	78	23	14	32	
Breed	Mixed	306	18	14	22	0.3659
	Spanish	37	27	13	41	
	Other countries	457	18	14	21	
Body weight	1.5–7.0	210	9	5	13	<0.0001
	7.1–15.0	189	18	13	23	
	15.2–27.0	177	19	13	25	
	27.5–73.0	216	27	21	33	

dog on a regular basis? and (iii) how many months and when in the year is your dog under insecticide treatment?

2.2. Blood sampling and *L. infantum* ELISA antibody analysis

Blood samples were taken from the cephalic vein into EDTA vacuum tubes, plasma recovered by centrifugation and used to detect *L. infantum* IgG antibodies with two commercial antibody tests: Ingezim Leishmania (Ingensa®, Spain) and Civtest Canis Leishmania (Hipra®, Spain, later commercialized by Esteve®, Spain as Leiscan). Both tests use crude immunodominant *L. infantum* antigens and conjugates include a specific canine IgG monoclonal in the Ingezim test and a generic protein A/HRPO in the Civtest. The validity of these tests has been assessed by manufacturers using the indirect immunofluorescence test (IFI) as the reference test. The Ingezim showed a 95% and 80% results agreement for 1/100 and 1/160 IFI cut-offs, respectively, and the estimated Civtest's sensitivity (Se) and specificity (Sp) was 98% and 96%, respectively. Moreover, the performance of both tests was recently assessed in experimentally infected dogs by Rodríguez-Cortés et al. (2013). Sp was 100% for both tests whilst Se were 98% for Leiscan and 78% for Ingezim.

Samples were done in duplicate and antibody optical densities (OD) were read in a spectrophotometer. The mean of both readings were used to classify samples as positive, negative or inconclusive, following kit instructions.

2.3. Spatial characterization of the dog's residential area

The dog's residential environment was characterized using a geographical information system (ArcGIS v.10, ESRI, Redlands, USA), and used with other dog variables, as covariates in the analysis of the relationship between the dogs CanL serological status and insecticide use. The approach taken was described in detail in a recent human Leishmaniosis study in Murcia (Pérez-Cutillas et al., 2015). Briefly, the dog's home address was used to obtain the geographical coordinates (<http://iderm.imida.es/>) and define a 500 m radius circular area around the point location from which environmental data was extracted (Tables 4 and 5). Ecological variables considered were the average monthly mean temperature, rainfall and relative humidity, obtained from an interpolation using point data from 45 regional weather stations (

Table 2
Canine Leishmaniosis seroprevalence (95% confidence intervals) according to dogs' insecticide usage. A cross-sectional study in dogs from Murcia, southeast Spain in autumn 2011.

Variable	Level	N	ELISA			P value
			% positive	95–	95+	
Regular use of insecticides	No	60	12	4	20	0.2218
	Yes	740	19	16	22	
No. of insecticides used in any one year	1 Product	522	20	17	24	0.6715
	2 Products	191	17	11	22	
	3 Products	10	10	0	29	
Insecticide application mode	Spot-on	334	19	14	23	0.1312
	Collar	192	20	14	25	
	Spot-on and collar	142	14	8	20	
	Spray	36	22	9	36	
Insecticidal product regularly used	Lotion/shampoo	18	39	16	61	0.3721
	Advantix spot-on ¹ only	159	20	14	26	
	Exspot spot-on ² only	40	25	12	38	
	Scalibor collar ³ only	127	22	15	9	
	Advantix spot-on and Scalibor collar	69	9	2	15	
	Advantix spot-on and other products	21	19	2	36	
	Exspot spot-on and Scalibor collar	25	16	2	30	
	Exspot spot-on and other products	16	31	9	54	
	Scalibor collar and other products	45	18	7	29	
	Other products ⁴	161	17	12	23	
Insecticidal product; annual treatment period (months)	Advantix spot-on only; 1–7	49	16	6	27	0.2002
	Advantix spot-on only; 8–12	106	21	13	28	
	Exspot spot-on only; 8–12	29	24	9	40	
	Advantix spot-on; 8–12 and Scalibor collar; 12	53	6	0	12	
	Exspot spot-on; 8–12 and Scalibor collar; 12	12	17	0	38	
	Scalibor collar only; 3–9	32	22	8	36	
Scalibor collar only; 12	88	22	13	30		

Advantix pipettes¹: 50% permethrin and 10% imidacloprid, Exspot pipettes²: 65% deltamethrin, Scalibor collar³: 4% deltamethrin, other products⁴: included pipettes and collars other than Scalibor collars and Advantix and Exspot pipettes as well as insecticide body shampoos and lotions.

Table 3
Canine Leishmaniosis seroprevalence (95% confidence intervals) according to dogs' home characteristics and sleeping place. A cross-sectional study in dogs from Murcia, southeast Spain in autumn 2011.

Variable	Level	N	ELISA			P value
			% positive	95–	95+	
Home type	Modern terraced	70	19	9	28	<0.0001
	Traditional terraced	141	16	10	22	
	Farmhouse	313	26	21	30	
	Detached house	143	16	10	22	
	Apartment	128	5	2	9	
Sleeping place	Indoors	433	12	9	15	<0.0001
	Outdoors	362	26	21	30	
Other dogs at home	No	340	16	12	19	0.1004
	Yes	461	20	17	24	
Cats at home	No	648	18	15	21	0.4269
	Yes	153	21	14	27	
Other animals at home	No	665	18	15	21	0.6741
	Yes	135	20	13	27	
No. people at home	0	60	30	18	42	0.1325
	1	67	16	8	25	
	2	239	15	10	19	
	3	163	18	12	24	
	4	179	21	15	27	
	≥5	63	19	9	29	

imida.es/), the mean ground altitude (Digital Elevation Model, NASA TERRA mission, <http://gdem.ersdac.jspacesystems.or.jp>) and the CORINE Land Cover use (1:100000) (<http://terrestrial.eionet.europa.eu/CLC2006>). In addition, individual dogs' residential environment were visually examined using orthoimages from WMS of IDERM and classified into one of eight habitat classes. They included the six previously defined human habitats (Pérez-Cutillas et al., 2015), i.e.: urban areas with a high or medium-low population density, periurban residential areas, and rural villages, hamlets and isolated farms, and two additional ones: industrial states and periurban orchard holdings (Table 4).

2.4. Statistical analysis

Seroprevalence was defined as the proportion of samples positive or inconclusive (ODs between testis negative and positive thresholds) to one or both ELISA tests. It was estimated globally and for specific explanatory variables (Tables 1–5). Seroprevalence and median log-transformed ODs across variable levels were compared using Yates-corrected Chi-squared test and the Kruskal–Wallis test, respectively (Kirwood and Sterne, 2003).

The relationship between the dogs serological status (seropositive or seronegative) and the insecticide product regularly used

Table 6
Estimates of the multivariable logistic regression model of the relationship between the dog's Canine Leishmaniosis serological status and dog demographic and residence variables. A cross-sectional study in dogs from Murcia, southeast Spain in autumn 2011 ($n = 588$ dogs).

Variable	Level	ELISA			P value
		Odd ratio	95–	95+	
Age (years)	1	1			
	2–3	1.21	0.55	2.69	0.6343
	4–6	1.99	0.93	4.26	0.0763
	7–15	2.78	1.32	5.85	0.0073
Bodyweight (kg)	1.5–7.0	1			
	7.1–15.0	2.91	1.36	6.25	0.0061
	15.2–27.0	2.00	0.85	4.70	0.1136
	27.5–73.0	3.82	1.73	8.40	0.0009
Sleeping place	Indoors	1			
	Outdoors	1.72	0.94	3.16	0.0784
Residential setting	Urban high density and industrial states	1			
	Urban low density	2.56	1.08	6.11	0.0337
	Periurban residential states	2.23	0.62	7.94	0.2172
	Periurban orchard holdings	1.92	0.81	4.56	0.1403
	Rural villages	2.50	0.71	8.80	0.1538
	Rural hamlets	3.56	1.02	12.48	0.0473
	Isolated ranches	3.58	1.09	11.82	0.0360
Altitude (m); Average March–July Ambient temperature (°C)	04–98; 19.1–19.5	1			
	04–98; 19.6–20.5	0.80	0.32	2.01	0.6362
	103–299; 18.6–19.0	0.32	0.06	1.77	0.1918
	103–299; 19.1–19.5	2.80	0.79	9.97	0.1116
	301–500; 18.6–19.0	4.48	1.87	10.75	0.0008
	503–878; 16.4–17.5	1.56	0.67	3.59	0.2998
Regular insecticide preventive Treatment	503–878; 17.6–18.5	1.15	0.45	2.95	0.7668
	No treatment	1			
	Advantix pipettes	2.14	0.73	6.27	0.1661
	Advantix pipettes ¹ and Scalibor collar ²	0.65	0.17	2.59	0.5458
	Advantix pipettes and other products ³	3.06	0.66	14.23	0.1542
	Exspot pipettes ⁴	3.62	1.02	12.84	0.0464
	Exspot pipettes and other products	14.45	2.15	97.26	0.0060
	Scalibor collar	1.83	0.62	5.36	0.2712
	Scalibor collar and Exspot pipettes	4.39	0.96	20.10	0.0568
	Scalibor collar and other products	1.94	0.47	8.05	0.3605
Other products	2.32	0.76	7.07	0.1370	

Insecticides: Advantix pipettes¹: 50% permethrin and 10% imidacloprid, Scalibor collar²: 4% deltamethrin, other products³: included pipettes and collars other than Scalibor collars and Advantix and Exspot pipettes as well as insecticide body shampoos and lotions, Exspot pipettes⁴: 65% deltamethrin.

Scalibor collars plus ExSpot pipettes, 20% ($n = 159$) in those with Advantix pipettes alone, 22% ($n = 127$) in dogs with Scalibor collars alone and 25% ($n = 40$) in those with Exspot pipettes alone ($p > 0.05$) (Table 2). Differences in seroprevalence between dogs using Scalibor collars plus Advantix pipettes and those using Scalibor collars or Exspot pipettes alone were significant ($p < 0.05$).

Treatment duration ranged from 1 to 12 months a year and was always administered from spring onwards when used for less than 12 months. The group of dogs with the lowest seroprevalence was 6% corresponding to those using Advantix pipettes and Scalibor collars for 8–12 months and 12 months a year, respectively (Table 2). Median ODs among seropositive dogs was similar for dogs using or not insecticides and for different insecticide brands and combinations ($p > 0.05$).

3.2. Univariate relationship between CanL seroprevalence and dogs residential environment

There were marked differences in CanL seroprevalence according to the dog's place of residence and it was 5% in dogs living in apartments compared to 26% among dogs living in a farm. Also, seroprevalence was 26% in dogs sleeping outside compared to 12% in those sleeping indoors ($p < 0.05$) (Table 3). Sharing a home with other dogs was also associated to higher prevalence although differences were not significant (Table 3).

The strong association between seroprevalence and the dog's residence is further exemplified when considering environmental variables in the dog's wider home surroundings. Seroprevalence was 9% in dogs in highly densely populated urban areas, 16% in

urban areas with a lower population density and in periurban residential states, 21% in rural villages, 23% in periurban orchard holdings, 28% in isolated ranches and 29% in rural hamlets ($p < 0.05$) (Table 4). Similarly, seroprevalence was 12% in areas dominated by artificial surfaces, 20% in green urbanized areas, 23% in predominantly natural areas and 15% in areas with a mixture of artificial and natural areas ($p < 0.05$) (Table 4).

CanL seroprevalence also varied depending on climatic variables. Areas with the highest prevalence had moderate or moderately high annual and seasonal temperatures, lower annual and autumn rainfall and lower relative humidity compared to areas with less seroprevalence, although differences in some cases were not significant (Table 5).

3.3. Relationship between CanL seroprevalence and insecticide use adjusted for dog demographic and environmental variables in the place of residence

The most parsimonious logistic regression model investigating the relationship between CanL seroprevalence and insecticides in dogs included age, body weight, sleeping place (outdoors or indoors), residential setting and a variable representing the combination between altitude and the average March–July temperature in the dog's residence (Table 6). There was no significant residual variation associated to the veterinary practice.

The model confirmed the lack of an association between dogs having insecticide treatment on a regular basis and reduced seroprevalence (Table 6). Among treated dogs, the combination of Advantix pipettes and Scalibor collars was associated with a sig-

nificantly ($p < 0.05$) or marginally significantly ($p < 0.10$) lower risk of being seropositive compared to dogs having these products alone or in combination with others (OR not shown). The model also indicated that seropositivity was greatest for dogs living in the 300–500 m altitude range with average March–July temperatures of 18.6–19.0 °C than dogs living below 100 m and with average March–July temperatures of 19.1–19.5 °C (Table 6).

4. Discussion

The percentage of dog owners using insecticides on a regular basis was over 90%, probably higher than that in the overall population in Murcia. Owners taking their dogs to the veterinarian and willing to have a CanL investigation would be more concerned about preventing infection and using insecticides. They were also likely to provide accurate information. However, it was considered unrealistic to ask them about the dog's life-long insecticide use and obtain accurate data. It's probable that owners of most young and middle age dogs used the same ectoparasite control program as recommended by their veterinarian, throughout their dog's life. This may have not been the case for older animals particularly those outliving the products life in the Spanish market (Advantix, Scalibor and Exspot were available since 2004, 2000 and 1997, respectively). Insecticidal products efficacy cannot be accurately estimated unless dogs are treated from the youngest age, before being exposed to sandflies and manufacturer's indications are strictly followed. Some dogs in the present study may have been infected before using the products claimed. It's also likely that products were not always used as recommended since owners may not be fully aware or find difficult to strictly adhere to manufacturer's indications. Study results however, provide a moment in time picture of CanL seroprevalence in dogs in relation to regular insecticide usage. They indicate that the availability of these products in an endemic area for a long time has not resulted in a significant reduction in CanL seroprevalence, although it was lowest in dogs using the combination of Scalibor collar and Advantix pipettes than either product alone or Exspot. This is an important finding with practical implications for the control of CanL. Experimental trials should be carried out to corroborate the efficacy of the double treatment with Advantix and Scalibor and to examine related dog health and safety issues. The latter is important because using the two antiparasitic formulations concomitantly may increase the risk of pyrethroid toxicity.

The intrinsic limitations associated to insecticidal products to prevent *Leishmania* spp. infection have been demonstrated experimentally. Advantix and Exspot's repellent efficacy against *P. perniciosus* is 87–99% during the first three weeks after application and is considerably reduced in the fourth and subsequent weeks (Miró et al., 2007; Molina et al., 2012). As mentioned previously, in controlled field trials Scalibor collars may only reduce the annual risk of seroconversion by 50% (Maroli et al., 2001; Foglia-Manzillo et al., 2006) hence, the risk of dogs becoming infected gradually increases year after year, possibly to the levels observed in this study. The dog's chance of becoming infected may be further increased if insecticides are not used throughout the entire sandfly season. Some owners in the present study used them for less than 8 months a year and this would not always cover the adult sandfly season in Murcia. Spot-on formulations may be washed off if dogs are soaked in water before the product is absorbed. Incorrect mechanical application of the product can also reduce its ability to prevent infection. Study findings highlight the need for veterinary practices to keep owners informed of the risks of vector exposure, and on the importance of proper use and limitations associated with insecticidal products to prevent CanL.

The reasons for the low seroprevalence among insecticide-free dogs are unclear. It is possibly a chance finding given the relatively small number of dogs in this group ($n = 60$). *L. infantum* seroprevalence is an indirect measure of the extent of CanL susceptibility (Baneth et al., 2008). Two serological tests were used here to increase diagnostic sensitivity. In contrast to the study by Rodríguez-Cortés et al. (2013), Ingezim detected a greater proportion of seropositives compared to Civtest. Variability in diagnostic tests performance could be related to differences in the number and origin of the dogs investigated. Our seroprevalence estimation when considering only Civtest positives as seropositive is similar to that in a previous study in dogs from periurban areas in Murcia (Chitimia et al., 2011). Given that both tests are highly specific (Rodríguez-Cortés et al., 2013), it should be advisable to employ both tests to detect a greater number of seropositives and investigate associated risk factors.

The study also provided useful baseline CanL epidemiologic data for Murcia, particularly for dogs visiting veterinary practices and using insecticidal treatments. Murcia is a typical southern Mediterranean *L. infantum* endemic area. Like in other studies in this area, seroprevalence increased with age except in oldest dogs and with bodyweight. The former could be associated to a greater death rate among seropositive dogs due to CanL and other concomitant infections, and the latter to bigger dogs being used as guard dogs and living permanently outside (Alvar et al., 2004). The dog's residential setting, altitude and temperature also had a major influence in seroprevalence, presumably by affecting sandfly abundance and exposure to infection. Studies in Murcia and nearby areas showed a similar relationship between leishmaniosis and altitude and temperature (Alonso et al., 2010; Pérez-Cutillas et al., 2015).

As expected, taking into account environmental and other dog-related variables is an essential part of population-based studies investigating the role of insecticides against vector-borne infections. They can partly overcome lack of accurate information on vector abundance, which is costly and time demanding to obtain. Moreover, the epidemiological information in this article can help design rational, cost-effective entomological studies. Future cross-sectional studies in Murcia would be required for a more accurate indication of the role played by insecticides in the dog on CanL seroprevalence in the region.

5. Conclusion

Using preventive anti-sandfly insecticide products on a regular basis appears to have no effect on CanL seroprevalence except possibly for the combination of Scalibor collars (deltamethrin 40 mg/g; MSD®) and Advantix pipettes (permethrin 500 mg/ml and imidacloprid 100 mg/ml; Bayer®). Stronger emphasis needs to be placed on informing dog owners of the best protocols to minimize sandfly infection and on identifying and targeting high prevalence areas to improve CanL control.

Conflict of interest

None of the authors of this paper has a financial or personal relationship with other people or organisations that could inappropriately influence or bias the content of the paper.

Acknowledgements

This study was funded by the Fundación Séneca de la Región de Murcia (Pr. Ref: 08714/PI/08), and the Spanish Ministry of Science and Innovation (Pr. Ref: AGL2013-46981-R) and the Instituto de Salud Carlos III within the Network of Tropical Diseases Research (RICET RD06/0021/1007). Authors of this paper are presently mem-

bers and receive support from COST Action TD1303 (European Network for Neglected Vectors and Vector-borne infections).

References

- Alonso, F., Giménez Font, P., Manchón, M., Ruiz De Ybáñez, R., Segovia, M., Berriatua, E., 2010. Geographical variation and factors associated to seroprevalence of Canine Leishmaniosis in an endemic Mediterranean area. *Zoonoses Public Health* 57, 318–328.
- Alvar, J., Cañavate, C., Molina, R., Moreno, J., Nieto, J., 2004. Canine leishmaniasis. *Adv. Parasitol.* 57, 1–88.
- Baneth, G., Koutinas, A.F., Solano-Gallego, L., Bourdeau, P., Ferrer, L., 2008. Canine leishmaniosis—new concepts and insights on an expanding zoonosis: part one. *Trends Parasitol.* 24, 324–330.
- Chitimia, L., Muñoz-García, C.I., Sánchez-Velasco, D., Lizana, V., Del Río, L., Murcia, L., Fisa, R., Riera, C., Giménez-Font, P., Jiménez-Montalbán, P., Martínez-Ramírez, A., Meseguer-Meseguer, J.M., García-Bacete, I., Sánchez-Isarria, M.A., Sanchis-Monsonís, G., García-Martínez, J.D., Vicente, V., Segovia, M., Berriatua, E., 2011. Cryptic Leishmaniosis by *Leishmania infantum*, a feature of canines only? A study of natural infection in wild rabbits, humans and dogs in southeastern Spain. *Vet. Parasitol.* 181, 12–16.
- de Ybáñez, R.R., del Río, L., Martínez-Carrasco, C., Segovia, M., Cox, J., Davies, C., Berriatua, E., 2009. Questionnaire survey on Canine Leishmaniosis in southeastern Spain. *Vet. Parasitol.* 164, 124–133.
- Ferroglio, E., Poggi, M., Trisciuglio, A., 2008. Evaluation of 65% permethrin spot-on and deltamethrin-impregnated collars for canine *Leishmania infantum* infection prevention. *Zoonoses Public Health* 55, 145–148.
- Foglia Manzillo, V., Oliva, G., Pagano, A., Manna, L., Maroli, M., Gradoni, L., 2006. Deltamethrin-impregnated collars for the control of canine leishmaniasis: evaluation of the protective effect and influence on the clinical outcome of *Leishmania infantum* infection in kennelled stray dogs. *Vet. Parasitol.* 142, 142–145.
- Killick-Kendrick, R., Killick-Kendrick, M., Focieux, C., Dereure, J., Puech, M.P., Cadiergues, M.C., 1997. Protection of dogs from bites of phlebotomine sandflies by deltamethrin collars for control of canine leishmaniasis. *Med. Vet. Entomol.* 11, 105–111.
- Kirwood, B.R., Sterne, J.A.C., 2003. *Essential Medical Statistics*, 2nd ed. Blackwell Publishing, Oxford, UK.
- Kleinbaum, D.G., Kupper, L.L., Muller, K.E., Nizam, A., 1998. *Applied Regression Analysis and Other Multivariable Methods*. Duxbury Press at Brooks/Cole Publishing Company.
- Maroli, M., Mizzon, V., Siragusa, C., D'Oorazi, A., Gradoni, L., 2001. Evidence for an impact on the incidence of canine leishmaniasis by the mass use of deltamethrin-impregnated dog collars in southern Italy. *Med. Vet. Entomol.* 15, 358–363.
- Martínez Ortega, E., Conesa Gallego, E., 1987. Fenología de los flebótomos del subgénero *Larrousius* (Dip. Psychodidae: Phlebotomus) en el sureste de la Península Ibérica. *Boletín Asoc. Esp. Entomol.* 11, 293–300.
- Mencke, N., Volf, P., Volfova, V., Stanneck, D., 2003. Repellent efficacy of a combination containing imidacloprid and permethrin against sand flies (*Phlebotomus papatasi*) in dogs. *Parasitol. Res.* 90 (Suppl. 3), S108–S111.
- Miró, G., Gálvez, R., Mateo, M., Montoya, A., Descalzo, M.A., Molina, R., 2007. Evaluation of the efficacy of a topically administered combination of imidacloprid and permethrin against *Phlebotomus perniciosus* in dog. *Vet. Parasitol.* 143, 375–379.
- Molina, R., Espinosa-Góngora, C., Gálvez, R., Montoya, A., Descalzo, M.A., Jiménez, M.I., Dado, D., Miró, G., 2012. Efficacy of 65% permethrin applied to dogs as a spot-on against *Phlebotomus perniciosus*. *Vet. Parasitol.* 187, 529–533.
- Otranto, D., Dantas-Torres, F., 2013. The prevention of canine leishmaniasis and its impact on public health. *Trends Parasitol.* 29, 339–345.
- Otranto, D., de Caprariis, D., Lia, R.P., Tarallo, V., Lorusso, V., Testini, G., Dantas-Torres, F., Latrofa, S., Diniz, P.P., Mencke, N., Maggi, R.G., Breitschwerdt, E., Capelli, G., Stanneck, D., 2010. Prevention of endemic canine vector-borne diseases using imidacloprid 10% and permethrin 50% in young dogs: a longitudinal field study. *Vet. Parasitol.* 172, 323–332.
- Otranto, D., Paradies, P., Lia, R.P., Latrofa, M.S., Testini, G., Cantacessi, C., Mencke, N., Galli, G., Capelli, G., Stanneck, D., 2007. Efficacy of a combination of 10% imidacloprid/50% permethrin for the prevention of leishmaniasis in kennelled dogs in an endemic area. *Vet. Parasitol.* 144, 270–278.
- Pérez-Cutillas, P., Goyena, E., Chitimia, L., De la Rúa, P., Bernal, L.J., Fisa, R., Riera, C., Iborra, A., Murcia, L., Segovia, M., Berriatua, E., 2015. Spatial distribution of human asymptomatic *Leishmania infantum* infection in southeast Spain: a study of environmental, demographic and social risk factors. *Acta Trop.* 146, 127–134.
- Reithinger, R., Coleman, P.G., Alexander, B., Vieira, E.P., Assis, G., Davies, C.R., 2004. Are insecticide-impregnated dog collars a feasible alternative to dog culling as a strategy for controlling canine visceral leishmaniasis in Brazil? *Int. J. Parasitol.* 34, 55–62.
- Rodríguez-Cortés, A., Ojeda, A., Todolí, F., Alberola, J., 2013. Performance of commercially available serological diagnostic tests to detect *Leishmania infantum* infection on experimentally infected dogs. *Vet. Parasitol.* 191, 363–366.
- Solano-Gallego, L., Morell, P., Arboix, M., Alberola, J., Ferrer, L., 2001. Prevalence of *Leishmania infantum* infection in dogs living in an area of canine leishmaniasis endemicity using PCR on several tissues and serology. *J. Clin. Microbiol.* 39 (2), 560–563.